

SEVEN GUAIANOLIDES FROM *EUPATORIUM CHINENSE*

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Key Word Index—*Eupatorium chinense*; Compositae; new sesquiterpene lactone; new guaianolides; hydroperoxides; ^1H and ^{13}C NMR; photo-oxygenation.

Abstract—Two allyl hydroperoxy guaianolide sesquiterpene lactones (peroxyeupahakonin-A and -B) and five new guaianolides (eupahakonin-A and -B, eupahakonenin-A and -B, and eupahakonesin) were isolated from *E. chinense* and characterized. The allyl hydroperoxy sesquiterpene lactones were characterized by spectral and chemical methods. They were prepared chemically by photosensitized oxygenation of eupahakonin-A.

INTRODUCTION

Previous studies on sesquiterpene lactones from the genus *Eupatorium* have yielded several germacranolides, heliangolides and guaianolides. For example, the four heliangolides eupalinin-A, -B, -C, and -D were isolated from *E. lindleyanum* [1]. Among these compounds, eupalinin-A showed significant inhibitory activity against KB cell culture. Eupachifolin-A, which is a *cis,cis*-4,5,9,10-germacradienolide possessing an α,β -unsaturated aldehyde group, and the related guaianolides the eupachifolins-B, -C, -D and -E were obtained from *E. chinense* var. *simplicifolium* [2]. Peroxy-sachalinin, sachalinin, and sachalin were obtained from *E. sachalinense*, along with a known compound, eupatoriopicrin, which is the main component of this plant [3]. In the present paper, we describe the isolation and structure determination of two new guaianolide hydroperoxides and five closely related guaianolides from *E. chinense* L. var. *hakonense* (Nakai) Kitam. The structure and stereochemistry of the new sesquiterpene lactones were determined by chemical transformations and extensive applications of ^1H and ^{13}C NMR spectrometry.

RESULTS AND DISCUSSION

Peroxyeupahakonin-A (**1**, $\text{C}_{20}\text{H}_{24}\text{O}_8$ mp 143–146° [α]_D –165°) and -B (**2**, $\text{C}_{20}\text{H}_{24}\text{O}_8$ mp 147–148° [α]_D +35.4°), and eupahakonin-A (**3**, $\text{C}_{20}\text{H}_{24}\text{O}_7$ oil [α]_D –146°) and -B (**4**, $\text{C}_{20}\text{H}_{24}\text{O}_7$ oil [α]_D +65.3°) were isolated by repeated column chromatography on Si gel and then separated by prep. TLC. The IR, UV, and ^1H NMR spectra of each of these guaianolides supported the presence of an α,β -unsaturated γ -lactone system and an ester group. Characteristic of this class of compounds a *trans* diaxial relationship between the protons at C-5–C-6 and C-6–C-7 was observed in the signal for H-6 which occurred as a well-defined one-proton doublet of doublets at δ 4.50 (J = 8.5, 11 Hz), δ 4.52 (J = 9, 10 Hz), δ 4.46 (J = 8.5, 11 Hz), and δ 4.46 (J = 9, 10 Hz) in **1**, **2**, **3**, and **4**, respectively (Table 1).

In the ^{13}C NMR spectra (Table 2) of **1** and **2**, seven oxygenated carbon atoms were observed; two car-

bonyls, two singly protonated, two doubly protonated, and one unprotonated. The peaks at δ 94.6 in **1** and δ 95.4 in **2** were uniquely characteristic of an allylic carbon (C-1) bearing a hydroperoxy group. Furthermore, both compounds gave a positive ferrous thiocyanate test [4]. Acetalization of **1** and **2** with Me_2CO and *p*-TsOH gave the acetonides **1a**: oil; IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 3550, 3300, 1775 and 1715; m/z 432 [M]⁺ and **2a**: oil; IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 3550, 3350, 1770, and 1715; m/z 432 [M]⁺, respectively. These results indicated that **1** and **2** contained the same ester group. In the ^1H NMR spectrum of **2**, the exocyclic methylene protons gave two singlets at δ 5.03 and 5.09, instead of the broad singlet due to C-10 vinyl methyl protons seen in **1**. Treatment of **1** with triphenylphosphine afforded an alcohol, which was identical in all respects with eupahakonin-A (**3**). Peroxyeupahakonin-B (**2**) was likewise converted to eupahakonin-B (**4**).

Alkaline hydrolysis of **3** with 10% KOH in aqueous dioxane, followed by acetylation with acetic anhydride and pyridine gave the acetate **3a**: IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 3600, 1765, 1740 and 1665; m/z 304 [M]⁺, which was also formed by the same reactions from **4**.

From the above physical data and experiments, it was clear that peroxyeupahakonin-A (**1**) and -B (**2**), as well as eupahakonin-A (**3**) and -B (**4**), were regioisomers of the double bond at C-10, and that eupahakonin-A (**3**) and -B (**4**) are reduction products of peroxyeupahakonin-A (**1**) and -B (**2**), respectively. The configuration of the hydroperoxy group at C-1 and the ester group at C-8 in **1** was determined as follows. The chemical shift of H-5 in **3** (δ 2.85) clearly indicated that the hydroxyl group was α , because of the δ 0.51 high field shift compared with that of H-5 in **1** (δ 3.36). On the other hand, α -orientation of H-8 was supported by the fact that 8.5% NOE was observed between H-7 and H-8. From the above physical data and experiments, the structures of peroxyeupahakonin-A, -B, eupahakonin-A, and -B were established as **1**, **2**, **3**, and **4**, respectively.

Eupahakonenin-A (**5**, $\text{C}_{20}\text{H}_{24}\text{O}_6$, oil, [α]_D –95°) and -B (**6**, $\text{C}_{20}\text{H}_{24}\text{O}_6$, oil, [α]_D +22°) were purified several times by prep. TLC. These new sesquiterpene lac-

Table 1. ^1H NMR data of compounds 1–7 (100 MHz, TMS as internal standard)

Compound	H-1	H-2	H-3	H-5	H-6	H-7	H-8	H-9	H-13	H-14	H-15	Miscellaneous
1†	—	2.58 m	5.40 s(br)	3.36 d (11)	4.50 dd (8.5; 11)	3.59 m	5.88 dd(br) (3; 6.3)	5.70 dd(br) (1; 6.3)	5.57 d(3) 6.11 d(3.4)	92 s(br)	1.92 s(br)	6.75 t(5.8, H-3'), 4.20 s(H-5'), 4.30 d(5.8, H-4')
1a†	—	2.36 m	5.40*	3.36 d(br) (11)	4.48 dd (8.5; 11)	3.52 m	5.88 dd(br) (3; 6.3)	5.70 d(br) (6.3)	5.57 d(3) 6.15 d(3.4)	1.92 s(br)	1.92 s(br)	6.78 m(H-3'), 4.24–4.42 m (H-4', 5'), 1.36 s(2 × Me)
2†	—	2.90*	5.60*	2.90*	4.52 dd (9; 10)	3.40 m	5.60*	2.49 dd(3.2; 14.5) 3.14 dd(3.9; 14.5)	5.65 d(3.2) 6.18 d(3.7)	5.03 s 5.09 s	1.84 s(br)	6.80 t(5.8, H-3'), 4.27 s(H-5'), 4.35 d(5.8, H-4')
2a†	—	2.58*	5.58 s*	2.85*	4.24*	3.60 m	5.58*	2.47 dd(3.2; 14) 3.12 dd(3.9; 14)	5.58 d(3) 6.10 d(3.7)	5.02 s 5.08 s	1.80 s(br)	6.79 m(H-3'), 4.24–4.54 m (H-4', 5'), 1.35 s(2 × Me)
3†	—	2.56 m	5.58 s(br)	2.85 s(br) (11)	4.46 dd (8.5; 11)	3.60 m	5.88 dd(br) (3; 6)	5.48 m	5.56 d(3.2) 6.10 d(3.8)	1.92 m	1.96 s(br)	6.77 t(5.8, H-3'), 4.22 s(H-5'), 4.35 d(5.8, H-4')
3a†	—	2.44 m	5.58*	2.77 d(br) (10.5)	4.35 dd (8.5; 10.5)	3.36 m	5.48*	2.32 dd(5; 14) 3.05 dd(5; 14)	5.51 d(3) 6.26 d(3.8)	4.94 s 5.16 s	1.89 s(br)	2.01 s(OAc)
4†	—	2.62 m	5.60*	2.86 d(br) (10)	4.46 dd (9; 10)	3.59 m	5.60 d*	2.36 dd(3.5; 14) 3.19 dd(3.9; 14)	5.58 d(3.5) 6.10 d(3.8)	4.91 s 5.07 s	1.81 s(br)	6.80 t(5.8, H-3'), 4.22 s(H-5'), 4.32 d(5.8, H-4')
5§	—	3.03 s*	5.57*	3.40 d(br) (10)	4.12 t (10)	3.06 m	5.60 m*	2.61 m	5.44 d(3) 6.15 d(3.5)	1.61 s	1.97 s(br)	6.80 t(5.8, H-3'), 4.31 s(H-5'), 4.38 d(5.8, H-4')
6§	3.17*	2.48 m	5.53*	2.81 t(br) (9)	4.48 dd (9; 10.5)	3.17*	5.60 m*	2.55 d (5)	5.52 d(3) (6.22 d(3.8)	4.86 s 4.98 s	1.84 s(br)	6.79 t(5.8, H-3'), 4.28 s(H-5') 4.35 d(5.8, H-4')
6a§	3.14*	2.42 m	5.52*	2.77 t(br) (9)	4.38 dd (9; 10.5)	2.96 m	4.32 m	2.50 d (5)	5.56 d(3) 6.33 d(3.8)	4.90 s 4.97 s	1.84 s(br)	—
6b§	3.27*	2.50*	5.59 m	2.50*	4.95 dd (2; 10.5)	—	—	3.37 d(br)(10) 3.50 d(br)(10)	2.18 d (2)	5.06 s	1.90 s(br)	—
6d§	3.24*	2.36 m	5.49 m	2.71*	4.03 dd (9; 11)	3.10 m	4.92 m	2.27 dd(6; 14.5) 2.70 dd(5; 14.5)	5.59 d(3) 6.18 d(3.8)	4.90 s 5.04 s	1.85 s(br)	2.14(OAc)
6e§	3.01*	2.40*	5.50 m	2.85*	4.06 dd (9; 11)	3.04 m	5.34 m	5.26 s(br)	5.66 d(3) 6.22 d(3.8)	1.81 s	1.92 s(br)	2.15(OAc)
6f§	3.27 q (8)	2.37 m	5.48 m	2.75*	3.99 dd (9; 11)	2.75*	3.90 m	2.26 dd(6; 14.5) 2.75 m	6.13 d(3) 6.23 d(3.8)	4.91 s 5.03 s	1.85 s(br)	—
7§	3.46 dd (6; 8)	5.64*	5.64*	2.69*	4.64 dd (9; 10)	3.23 m	5.64*	2.69*	5.54 d(3) 6.31 d(3.5)	5.06 s 5.11 s	2.02 s	6.85 t(5.8, H-3'), 4.32 s(H-5'), 4.39 d(5.8, H-4'), 2.0 s(OAc)
7a§	3.44 dd (6; 8)	5.57*	5.67 s(br)	2.60*	4.57 dd (9; 10)	3.20 m	5.57*	2.60*	5.49 d(3) 6.31 d(3.5)	5.03 s 5.09 s	2.01 s	6.75 m(H-3'), 4.35; 4.44 m(H-4', 5'), 2.01 s(OAc), 1.44 s(2 × Me)
7b§	3.42 dd (6; 8)	5.51*	5.62 s(br)	2.50*	4.59 dd (9; 10)	3.13 m	5.51*	2.50*	5.48 d(3) 6.28 d(3.5)	5.00 s 5.06 s	1.98 s	1.97 s(2 × OAc)
7c§	3.42 dd (6; 8)	5.58*	5.64 s(br)	2.72*	5.59 dd (8.5; 11)	3.18 m	5.56*	2.53 dd(8; 14) 2.79 dd(6.5; 14)	5.48 d(3) 6.26 d(3.8)	5.03 s 5.08 s	1.99 s	6.67 t(5.8, H-3'), 4.32 d(5.8, H-4'), 4.35 d(5.8, H-4')

Figures in parentheses are coupling constants in Hz.

*Signal partially obscured or superimposed.

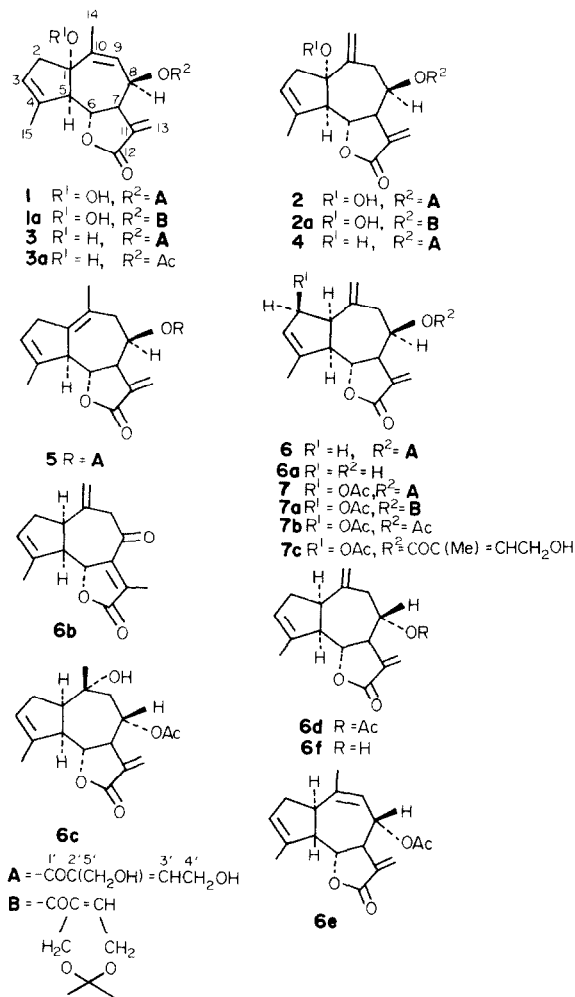
† $\text{d}_6\text{-Me}_2\text{CO}$.§ CD_3OD .§ CDCl_3 .

Table 2. ^{13}C NMR data of compounds 1–7 (25.05 MHz, TMS as internal standard)*

	1 (d_6 -Me $_2$ CO)	2 C $_3$ D $_3$ N)	3 (d_6 -Me $_2$ CO)	4 (d_6 -Me $_2$ CO)	5 (CDCl $_3$)	6 (CDCl $_3$)	7 (CDCl $_3$)	6d (CDCl $_3$)	6e (CDCl $_3$)
1	94.6 s	95.4 s	83.4 s	84.5 s	125.2 s	47.6 d	50.6 d	47.7 d	47.5 d
2	44.2 t	43.3 t	47.7 t	47.8 t	37.9 t	37.3 t	80.2 d	36.6 t	37.9 t
3	123.4 d	125.6 d	123.6 d	125.4 d	125.4 d	126.6 d	126.1 d	126.3 d	125.6 d
4	142.7 s†	144.5 s†	145.5 s†	147.0 s†	139.9 s†	139.6 s†	147.9 s†	140.9 s†	138.2 s†
5	60.7 d	59.8 d	66.4 d	64.8 d	56.4 d	56.4 d	55.8 d	56.7 d	55.1 d
6	78.9 d	79.3 d	79.0 d	79.7 d	79.9 d	80.0 d	80.0 d	80.2 d	79.5 d
7	48.2 d	48.5 d	48.2 d	47.8 d	55.3 d	48.5 d	47.9 d	49.3 d	46.8 d
8	67.9 d	68.0 d	67.8 d	68.7 d	66.7 d	68.2 d	68.3 d	74.2 d	73.1 d
9	122.8 d	37.1 t	119.6 d	36.7 t	37.6 t	40.3 t	39.0 t	38.4 t	121.6 d
10	141.7 s†	137.5 s†	142.2 s†	139.2 s†	136.7 s†	143.1 s†	138.7 s†	142.6 s†	142.7 s†
11	135.6 s	135.5 s	135.9 s	136.3 s	135.2 s	134.4 s†	133.7 s†	137.0 s†	136.6 s†
12	169.0 s	169.3 s	169.0 s	169.1 s	169.1 s	169.4 s	169.1 s	169.6 s	169.7 s
13	121.6 t	121.5 t	121.2 t	121.1 t	119.5 t	122.0 t	122.5 t	122.0 t	122.9 t
14	24.4 q	118.1 t	24.9 q	115.8 t	24.1 q	116.4 t	120.1 t	116.8 t	27.7 q
15	17.8 q	16.9 q	17.9 q	17.1 q	17.7 q	16.7 q	17.2 q	17.2 q	17.6 q
1'	166.4 s	166.4 s	166.5 s	166.2 s	165.9 s	165.8 s	165.8 s	—	—
2'	131.8 s	132.1 s	132.0 s	132.2 s	131.1 s	131.3 s	131.1 s	—	—
3'	145.9 d	145.8 d	145.9 d	145.9 d	144.1 d	143.9 d	144.0 d	—	—
4'	59.2 t	58.7 t	59.2 t	59.1 t	58.7 t	58.8 t	58.8 t	—	—
5'	56.9 t	56.3 t	57.0 t	56.9 t	56.7 t	56.9 t	56.8 t	—	—
Ac(C=O)	—	—	—	—	—	—	170.1 s	168.9 s	169.0 s
Ac(CH $_3$)	—	—	—	—	—	—	21.3 q	21.2 q	21.2 q

* Assignment established by single frequency off-resonance decoupling.

† Assignments may be interchanged.



tones were similar to eupahakonin-B (4) on the basis of the IR, UV, and mass spectral data, except for the absence of a *tert*-hydroxyl group. Extensive double irradiation 1H NMR experiments of **5** and **6** established the steric arrangements of the portion consisting of C-6–C-7. Further, the 1H NMR spectrum of **5** was similar to that of **6**, except for the presence of the vinyl-methyl group and the absence of exocyclic methylene protons, indicating that these compounds might be regioisomers of the double bond at C-10. The stereochemistry at C-1 and the ester group at C-8 in **6** was determined as follows. Hydrolysis of **6** with 10% KOH in aqueous dioxane, followed by oxidation of the resulting alcohol (**6a**, mp 144–146°, m/z 246 $[M]^+$) with pyridinium dichromate

[5] gave the butenolide **6b** (oil, mass spectrum m/z 244 $[M]^+$).

Dehydration of cumambrin-A (**6c**), isolated from *Crysanthemum ornatum* var. *spontaneum* [6], with $SOCl_2$ in pyridine gave **6d** (mp 89–91°) and **6e** (mp 101–102.5°). On hydrolysis with 10% KOH in aqueous dioxane, the acetate **6d** afforded the alcohol **6f** (mp 125–130° (decomp.), m/z 246 $[M]^+$), the 1H and ^{13}C NMR spectra of which were almost superimposable on those of **6a** except for the configuration at C-8. Oxidation of **6f** with pyridinium dichromate gave a compound, which was identical with the butenolide **6b**. Hence, the stereochemistry of H-1 and H-8 of eupahakonin-B (**6**) was α . From the above results, the structures of eupahakonin-A and -B were established as **5** and **6**, respectively.

Furthermore, photo-oxygenation of **5** in MeOH with visible light and methylene blue as sensitizer afforded peroxyeupahakonin-A (**1**) and -B (**2**), thereby confirming the absolute structure of (**5**).

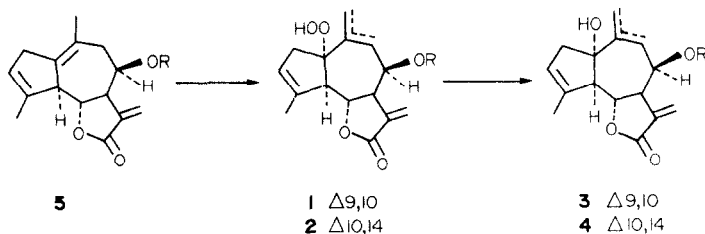
Eupahakonesin (**7**, oil, $[\alpha]_D -49.5^\circ$) gave rise to a molecular ion at m/z 418, in agreement with molecular formula $C_{22}H_{26}O_8$. Treatment of **7** with *p*-TsOH in acetone afforded the acetonide **7a** (oil, m/z 458 $[M]^+$). In addition, alkaline hydrolysis of **7** with 10% Na_2CO_3 in aqueous dioxane, followed by acetylation gave the diacetate **7b** (oil, m/z 346 $[M]^+$). From the above experiments and physical data (see Experimental and Tables 1 and 2), this new guaianolide had the same absolute stereostructure as eupahakonin-B (**6**) except for the additional presence of an acetoxy group [IR $\nu_{max}^{CHCl_3} cm^{-1}$: 3450 (OH), 1770 (γ -lactone), 1735 (OAc and ester), and 1660 (C=C)]; 1H NMR (Table 1) and ^{13}C NMR (Table 2)]. Further, the 1H NMR spectrum of **7** was very similar to that of eupachifolin-C (**7c**) which was isolated from *E. chinense* var. *simplicifolium* [2], except for the ester group. Hence, the structure of eupahakonesin was represented by **7**.

The co-occurrence of peroxyeupahakonin-A (**1**) and -B (**2**), and eupahakonin-A (**3**), -B (**4**), and eupahakonesin (**5**) in this plant is strongly indicative of a biosynthetic interrelationship between these compounds. A sequence of biogenetically feasible steps is shown in Scheme 1.

EXPERIMENTAL

Mps are uncorr. 1H and ^{13}C NMR spectra were measured at 100 and 25.05 MHz, respectively. MS, direct inlet, 70 eV. $[\alpha]_D$, UV, CD, and ORD in MeOH. TLC and prep. TLC: precoated Si gel plates (0.25 mm) F_{254} ; spots detected by UV light (254 nm) and by spraying with 1% $CeSO_4$ –10% H_2SO_4 .

Extraction and separation. The MeOH extract of a fresh whole plant (680 g) of *E. chinense* var. *hakonense* collected



Scheme 1. Biosynthetic relationships of compounds 1–5.

at Toyone, Aichi prefecture, Aug. 1979, was divided into the *n*-hexane- (10.9 g), CHCl_3 - (4.9 g) and EtOAc-soluble fractions (3.1 g). The crude CHCl_3 extract was chromatographed on Si gel using a CHCl_3 - Me_2CO gradient (6:1 starting mixture). 50-ml fractions were collected: fraction 1 (148.5 mg; CHCl_3 - Me_2CO , 6:1), fraction 2 (336.8 mg; CHCl_3 - Me_2CO , 6:1), fraction 3 (556 mg; CHCl_3 - Me_2CO , 4:1) and fraction 4 (148.5 mg; CHCl_3 - Me_2CO , 2:1). Fraction 2 was purified on a smaller Si gel column using CHCl_3 -MeOH (14:1) as solvent. The main fractions (220 mg) showed a single spot; prep. TLC (10% AgNO_3 -Si gel, C_6H_6 -*iso*-PrOH, 6:1) gave eupahakonenin-A (5) (66.7 mg) and -B (6) (76.6 mg). Fraction 3 was rechromatographed on Si gel using a CHCl_3 -MeOH gradient to give the less polar fractions (232.5 mg) and the more polar fractions (108.7 mg), respectively. The less polar fraction yielded eupahakonesin (7) (81 mg), while the more polar fraction was purified by prep. TLC (10% AgNO_3 -Si gel; C_6H_6 -*iso*-PrOH, 6:1) to afford peroxyeupahakonin-A (1) (41.4 mg) and -B (2) (48.4 mg), respectively. Fraction 4 was rechromatographed on Si gel using a CHCl_3 -MeOH gradient to give two major products (32.3 mg) which ran together on TLC. These were purified several times by prep. TLC (10% AgNO_3 -Si gel; C_6H_6 - CHCl_3 - Me_2CO , 3:2:2) to give pure eupahakonin-A (3) (7 mg) and -B (4) (3.3 mg), respectively.

Peroxyeupahakonin-A (1). $\text{C}_{20}\text{H}_{24}\text{O}_8$; mp 143–146° (amorph., Et_2O -EtOAc); $[\alpha]_D^{26} -165^\circ$ (MeOH; $c = 0.21$); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 209 (ϵ 16,491); IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3450, 1760, 1715, 1660; CD curve: $[\theta]_{255} -3378$, $[\theta]_{295} 0$; ORD (c 0.19): negative plane curve.

Peroxyeupahakonin-B (2). $\text{C}_{20}\text{H}_{24}\text{O}_8$; mp 147–148° (colourless needles, Me_2CO); $[\alpha]_D^{26} +35.4^\circ$ (MeOH; c 0.18); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 210 (ϵ 16000); IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3420–3250, 1765, 1715, 1655; CD curve: $[\theta]_{222} 0$, $[\theta]_{228} +1664$, $[\theta]_{237} 0$, $[\theta]_{260} -1821$, $[\theta]_{290} 0$; ORD (c 0.2): $[\phi]_{360}^{\text{T}} +22 \times 10^2$, $[\phi]_{335}^{\text{P}} +28 \times 10^2$, $[\phi]_{310}^{\text{T}} +46.2 \times 10^2$, $[\phi]_{280}^{\text{T}} +76.4 \times 10^2$.

Eupahakonin-A (3). $\text{C}_{20}\text{H}_{24}\text{O}_7$; colourless oil; $[\alpha]_D^{25} -146^\circ$ (c 0.2, MeOH); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 210 (ϵ 16200), 300 (ϵ 243); IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3400–3450, 1760, 1710, 1660; CD curve: $[\theta]_{247} -2575$, $[\theta]_{257} -3277$, $[\theta]_{292} 0$; ORD (c 0.2): negative plane curve.

Eupahakonin-B (4). $\text{C}_{20}\text{H}_{24}\text{O}_7$; colourless oil; $[\alpha]_D^{25} +65.3^\circ$ (MeOH; c 0.2); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 212 (ϵ 14395); IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3430, 1760, 1715, 1660; CD curve: $[\theta]_{210} -11,660$, $[\theta]_{224} 0$, $[\theta]_{229} +1167$, $[\theta]_{234} 0$, $[\theta]_{246} -1944$, $[\theta]_{259} -2334$, $[\theta]_{288} 0$; ORD (c 0.2): $[\phi]_{361}^{\text{T}} 10.8 \times 10^2$, $[\phi]_{335}^{\text{P}} +15 \times 10^2$, $[\phi]_{311}^{\text{T}} 12.2 \times 10^2$, $[\phi]_{280}^{\text{T}} +16 \times 10^2$.

Eupahakonenin-A (5). $\text{C}_{20}\text{H}_{24}\text{O}_6$; colourless oil; $[\alpha]_D^{25} -95^\circ$ (MeOH; c 0.2); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 210 (ϵ 16146); IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 3450, 1775, 1715, 1655; MS m/z : 360 $[\text{M}]^+$, 342, 246, 228; CD curve: $[\theta]_{222} -40230$, $[\theta]_{250} -8382$, $[\theta]_{267} 0$, $[\theta]_{278} +587$, $[\theta]_{295} 0$; ORD (c 0.19): negative plane curve.

Eupahakonenin-B (6). $\text{C}_{20}\text{H}_{24}\text{O}_6$; colourless oil; $[\alpha]_D^{25} +22^\circ$ (MeOH; c 0.19); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 210 (ϵ 15692); IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 3450, 1770, 1715, 1645; MS m/z : 360 $[\text{M}]^+$, 342, 246, 228; CD curve: $[\theta]_{216} -10903$, $[\theta]_{230} 0$, $[\theta]_{260} -2019$, $[\theta]_{285} 0$; ORD (c 0.2): $[\phi]_{361}^{\text{T}} +8.9 \times 10^2$, $[\phi]_{335}^{\text{P}} +12.8 \times 10^2$, $[\phi]_{310}^{\text{T}} +18.3 \times 10^2$, $[\phi]_{280}^{\text{P}} +13.5 \times 10^2$.

Eupahakonesin (7). $\text{C}_{22}\text{H}_{26}\text{O}_8$; colourless oil; $[\alpha]_D^{20} -49.5^\circ$ (MeOH; c 0.2); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 211.5 (ϵ 14209); IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 3450, 1770, 1735, 1660; MS m/z : 418 $[\text{M}]^+$, 400, 376, 358, 340, 304, 262, 244, 226; CD curve: $[\theta]_{244} -7511$.

Acetalization of 1. To a soln of 8.6 mg 1 in 2 ml Me_2CO was added 5 mg *p*-TsOH, and the soln stirred for 4 hr at room temp. The soln was neutralized with NaHCO_3 and poured into ice- H_2O . The CH_2Cl_2 extract was washed with H_2O , dried over Na_2SO_4 , and then evapd. The crude product

was purified through a small Si gel column to give 1a (7.3 mg): colourless oil; IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 3550, 3300, 1775, 1715, 1660; MS m/z 432 $[\text{M}]^+$, 417, 401, 374, 358, 340, 264, 260, 244, 226.

Treatment of 1 with triphenylphosphine. A MeOH soln of 1 (23.9 mg) was treated with triphenylphosphine (24.4 mg) at room temp. for 1 hr. Removal of the solvent gave a gummy material which was purified on Si gel column to afford a pure compound (17.2 mg) which was identical in all respects with eupahakonin-A (3).

Acetalization of 2. A Me_2CO soln of 2 (12.4 mg) was treated with *p*-TsOH (5 mg) with stirring for 3 hr at room temp. Work-up as 1a gave 2a (10.9 mg): colourless oil; IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 3550, 3350, 1770, 1715, 1662; MS m/z : 432 $[\text{M}]^+$, 416, 401, 374, 358, 340, 260, 244, 226.

Treatment of 2 with triphenylphosphine. 2 (28.9 mg) was treated with triphenylphosphine in the same manner as 1 to afford eupahakonin-B (4) (16 mg).

Hydrolysis of 3. To a soln of 3 (17.2 mg) in dioxane (2 ml) was added 10% KOH soln (4 ml). The reaction mixture was stirred for 6 hr at room temp. and after the usual work-up, the crude product was purified by prep. TLC to give a pure sesquiterpene lactone (1.7 mg) as a colourless oil. The lactone was acetylated with Ac_2O (0.5 ml) and pyridine (1 ml) for 24 hr at room temp. The usual work-up afforded the monoacetate 3a (0.6 mg): colourless oil; UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 213 (ϵ 4023); IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 3600, 1765, 1740, 1665, 1635; CD curve: $[\theta]_{217} -10,462$, $[\theta]_{246} -666$, $[\theta]_{260} -913$, $[\theta]_{295} 0$; MS m/z : 304 $[\text{M}]^+$, 262, 244. 3a was also obtained from 4 by the same set of reactions.

Hydrolysis of 6. A soln of 6 (54.4 mg) in dioxane (5 ml) was treated with 10% KOH-dioxane (3:1, 5 ml) at room temp. for 3 hr. After the usual work-up, the crude product was separated by prep. TLC (CHCl_3 -MeOH, 9:1) to give 6a: colourless needles (28.5 mg), mp 144–146° (Et_2O); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 208 (ϵ 8764); IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 3620, 1768, 1670, 1645; MS m/z : 246 $[\text{M}]^+$, 228; CD curve: $[\theta]_{210} 0$, $[\theta]_{220} -20048$, $[\theta]_{254} -802$, $[\theta]_{260} -859$, $[\theta]_{290} 0$.

Oxidation of 6a. 6a (13.2 mg) was stirred with pyridinium dichromate (60.3 mg) in dry CH_2Cl_2 (5 ml) at room temp. for 7 hr. After addition of Et_2O . The reaction mixture and filtered through a Celite pad and the filtrate concd *in vacuo*. The crude product was purified by prep. TLC to give the butenolide 6d (0.6 mg): colourless oil; UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (ϵ): 206 (4502), 243 (5348); IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 1765, 1695, 1645; MS m/z : 244 $[\text{M}]^+$.

Dehydration of 6c. To a pyridine soln of 6c (100 mg) was added SOCl_2 (116.3 mg) at 0° and the mixture allowed to stand for 1.5 hr. Work-up as usual gave a crude oil which showed two spots on TLC. It was purified by prep. TLC (*n*-hexane-EtOAc, 9:1) to afford 6d (16.8 mg) and 6e (35.5 mg). 6d: colourless needles, mp 89–91° (petrol- Et_2O); $[\alpha]_D^{23} +139.5^\circ$ (CHCl_3 ; c 0.2); IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 1770, 1740, 1665, 1645; MS m/z : 288 $[\text{M}]^+$, 228. 6e: colourless needles, mp 101–102.5° (*iso*-Pr $_2\text{O}$ - Et_2O); $[\alpha]_D^{23} +75^\circ$ (CHCl_3 ; c 0.2); IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 1775, 1740, 1665; MS m/z : 288 $[\text{M}]^+$, 244, 228.

Hydrolysis of 6d. 6d (16 mg) was treated with 10% KOH in dioxane under the same conditions as 6 to give the alcohol 6f (6 mg): colourless needles, mp 125–130° (decomp. Et_2O); IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 3600, 1765, 1665, 1640; MS m/z : 246 $[\text{M}]^+$, 228.

Oxidation of 6f. 6f (6 mg) was oxidized with pyridinium dichromate in dry CH_2Cl_2 under the same condition as 6a to afford a sesquiterpene lactone (0.3 mg) whose physical and spectral data were identical to those of 6b.

Photo-oxygenation of 5 to give 1 and 2. A soln of 5

(55.1 mg) and methylene blue (1.14 mg) in MeOH (13 ml) was irradiated internally with a high-pressure Hg lamp (Riko UVL-400P with Pyrex filter) in a H₂O-cooled immersion apparatus through which O₂ was circulated for 25 min. Removal of the solvent gave an oil which was purified by column chromatography on Si gel (CHCl₃-MeOH, 9:1). The fraction which was positive to the ferrous thiocyanate test was purified by repeated prep. TLC (10% AgNO₃-Si gel, C₆H₆-*iso*-PrOH, 6:1) to afford peroxyeupahakonin-A (**1**) (3.9 mg) and -B (**2**) (5.8 mg).

Acetalization of 7. Me₂CO soln of **7** (4.9 mg) was treated with *p*-TsOH under the same condition as **1** to afford the acetonide **7a** (5.1 mg): colourless oil; IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 1770, 1720-1735, 1660; MS *m/z*: 458 [M]⁺, 416, 400, 398, 358, 340, 322, 244, 227.

Hydrolysis of 7. **7** (81.8 mg) was treated with 10% Na₂CO₃-dioxane (3:1, 10 ml) soln at room temp. for 12 hr. Work-up as usual gave an oil which was purified by prep. TLC to give a pure alcohol, which was then acetylated by the usual manner to afford the diacetate **7b** (3.3 mg): colour-

less oil; IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 1765, 1735, 1660; MS *m/z*: 346 [M]⁺, 304, 286, 261, 244, 226.

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